

Shinning light on neuronal circuits

João Peça, jpeca@cnc.uc.pt



What controls behavior?



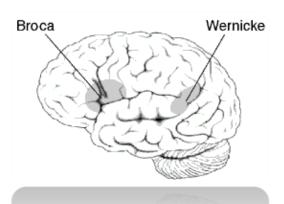


What is the Hegemonikon

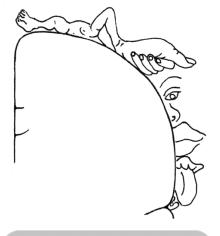
Neuronal circuits



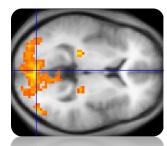
Joseph Gall 1810-



Paul Broca, 1861



Wilder Penfield, 1950





Ramachandran Damásio Logothetis

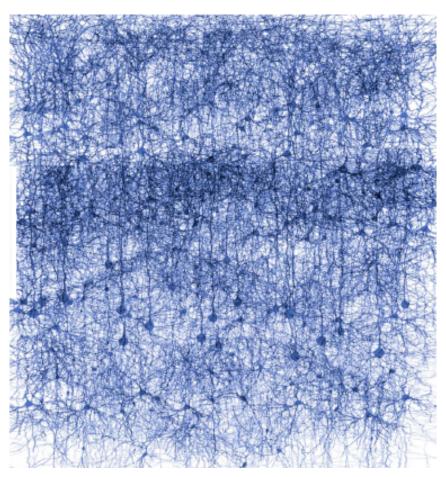
. . .

Crick 1979

it. For example, a method that would make it possible to inject one neuron with a substance that would then clearly stain all the neurons connected to it, and no others, would be invaluable. So would a method by which all neurons of just one type could be inactivated, leaving the others more or less unaltered.

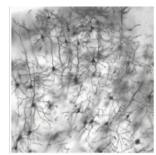
Visualization Control

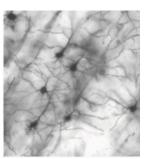
Complexity of the brain

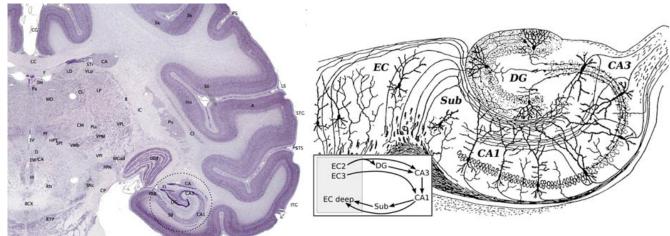


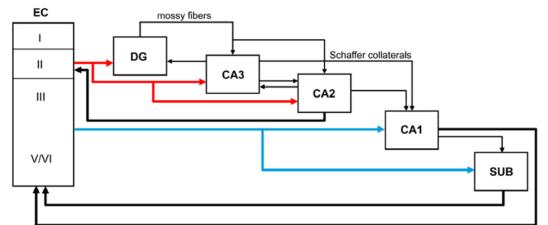
Only 1 in 100 neurons is shown here H. Markram, Blue Brain project

From Golgi to Diagrams









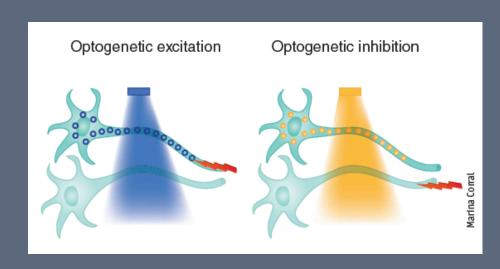
Circuit reconstruction allows for detailed on information flow and architecture

Llorens-Martin, M., et al. (2014). Front Neuroanat 8: 38.

Cartography of the brain

Mapping <u>functional circuitry</u> in the nervous system is a major goal for both cellular and system neuroscience

- Many methods have been developed to map functional connectivity of neural circuitry:
 - Electrophysiological stimulation
 - Glutamate uncaging
 - Ligand-gated ion channel coupled with photo-uncaging (P2X receptor)
- An ideal technique would require
 - Rapid photoactivation
 - Precise temporal control
 - High spatial resolution
 - Genetic targetability



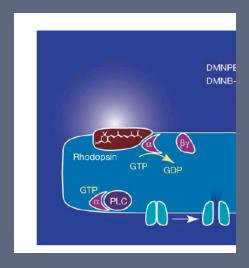
"3-step" optogenetics

Coexpression of the Drosophila photoreceptor genes encoding <u>arrestin-2</u>, <u>rhodopsin</u>, and the <u>α subunit of the cognate heterotrimeric G protein</u>—an explosive combination we term "chARGe"—sensitizes generalist vertebrate neurons

Neuron, Vol. 33, 15-22, January 3, 2002, Copyright @2002 by Cell Press

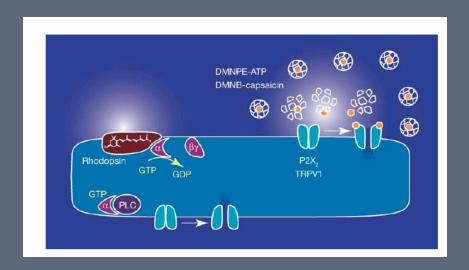
Selective Photostimulation of Genetically ChARGed Neurons

Neurotechnique



"2-step" optogenetics

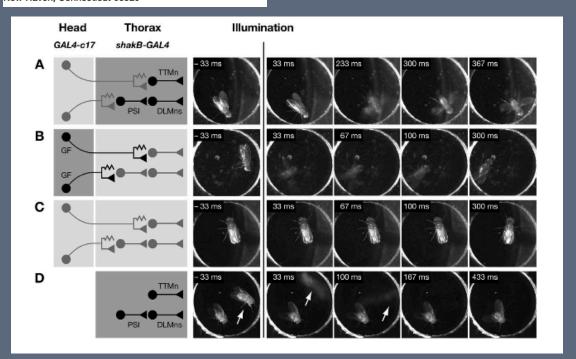
Because the fly genome lacks <u>purinoceptor</u> sequences (Littleton and Ganetzky, 2000), photoreleased ATP is expected to act selectively on the genetically designated targets.

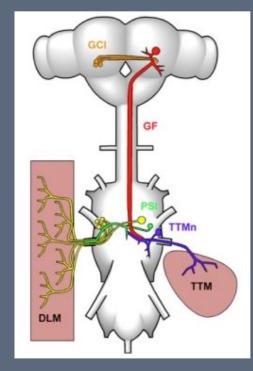


"2-step" optogenetics in action

Remote Control of Behavior through Genetically Targeted Photostimulation of Neurons

Susana Q. Lima and Gero Miesenböck* Department of Cell Biology Yale University School of Medicine 333 Cedar Street New Haven, Connecticut 06520

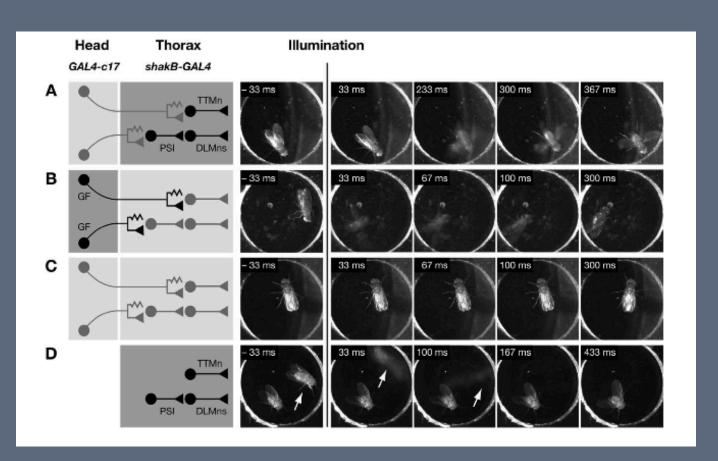




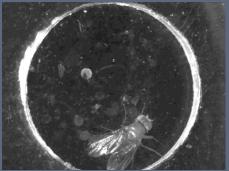
http://www.sussex.ac.uk/lifesci/bac onlab/research/giantfibre

Giant fiber (GF) neurons in the brain Thoracic ganglion contains TTMn, PSI, and DLMns

"2-step" optogenetics

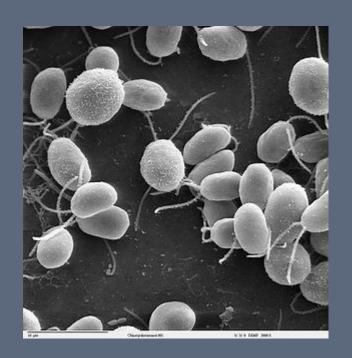


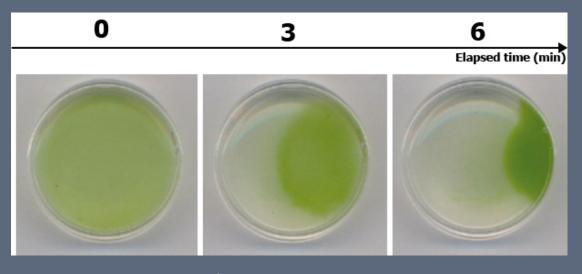




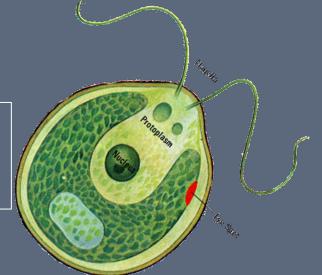


"Algae Vision"





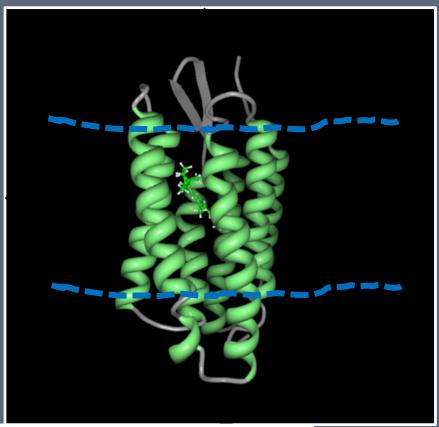
Chlamydomonas reinhardtii microalgae shows phototaxic behavior

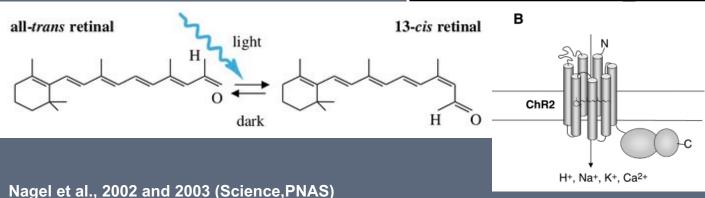


Channelrhodopsin-1: A Light-Gated Proton Channel in Green Algae

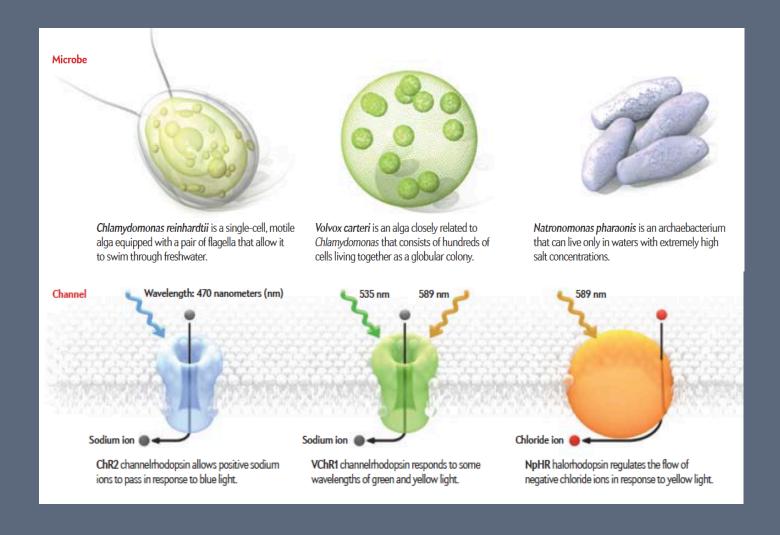
Georg Nagel, 1* Doris Ollig, 1 Markus Fuhrmann, 2 Suneel Kateriya, 2 Anna Maria Musti, 3 Ernst Bamberg, 1 Peter Hegemann 2

Phototaxis and photophobic responses of green algae are mediated by rhodopsins with microbial-type chromophores. We report a complementary DNA sequence in the green alga *Chlamydomonas reinhardtii* that encodes a microbial opsin-related protein, which we term Channelopsin-1. The hydrophobic core region of the protein shows homology to the light-activated proton pump bacteriorhodopsin. Expression of Channelopsin-1, or only the hydrophobic core, in *Xenopus laevis* oocytes in the presence of all-*trans* retinal produces a light-gated conductance that shows characteristics of a channel selectively permeable for protons. We suggest that Channelrhodopsins are involved in phototaxis of green algae.





Other channels and pumps

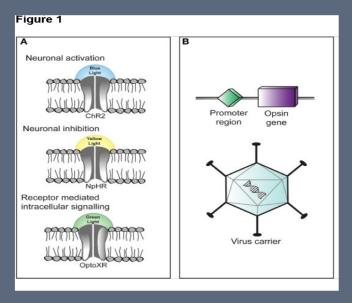


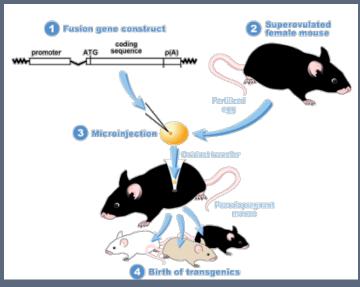
How to start optogenetics experiments?

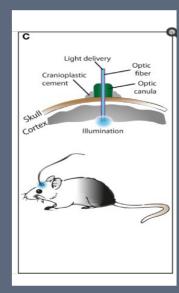
Gene therapy based

Transgene based

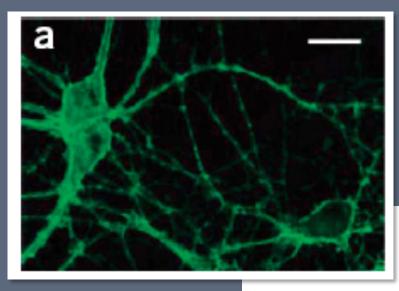
Activation

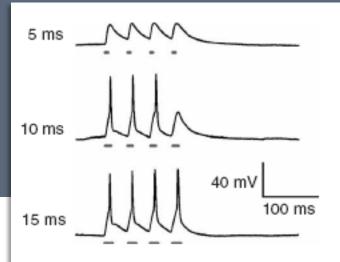




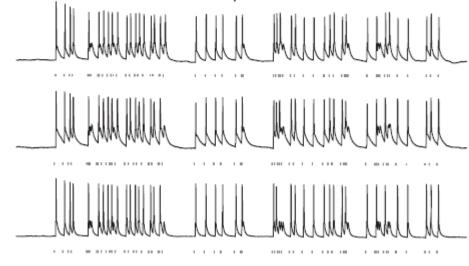


Typical in vitro result

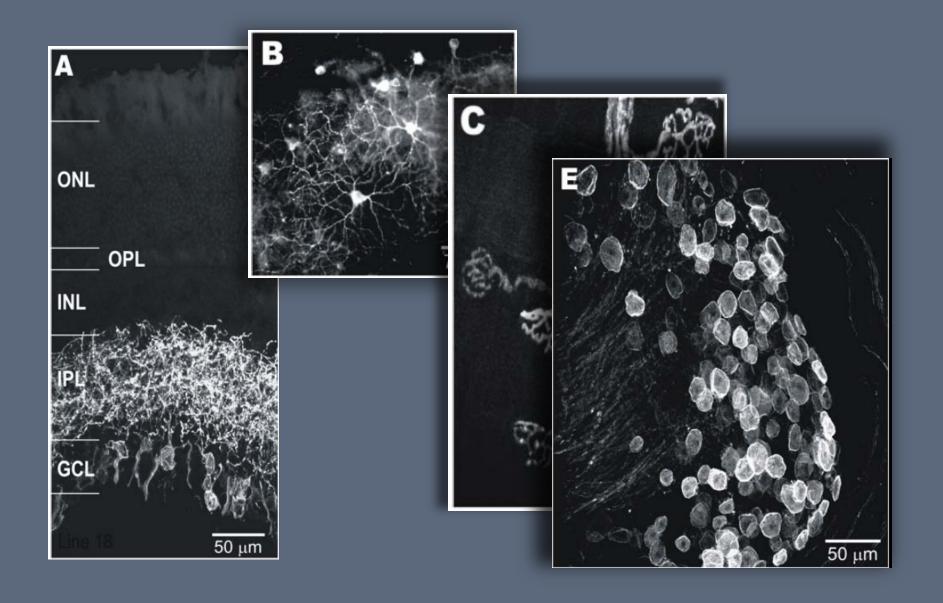




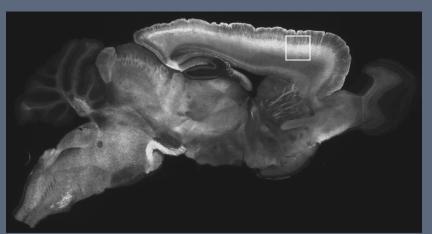
Three different neurons: same pulse series



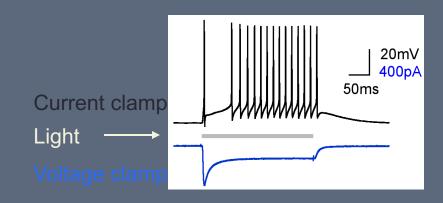
Characterization of ChR2 transgenic lines



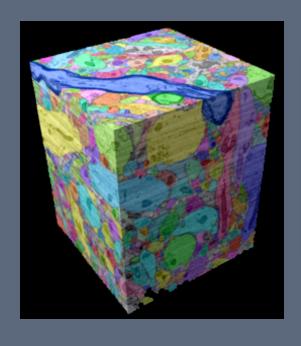
ChR2-mice positive neurons can be activated by light without exogenous retinal

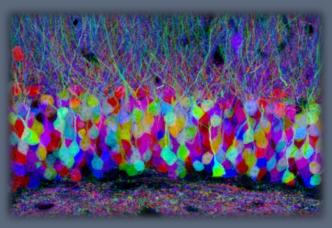


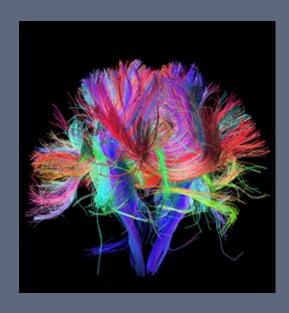




ChR2-assisted dissection of Microcircuits

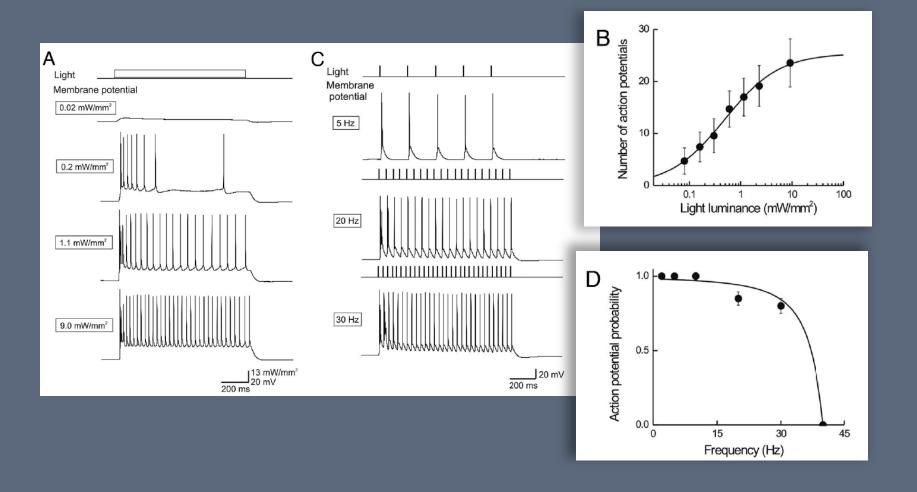




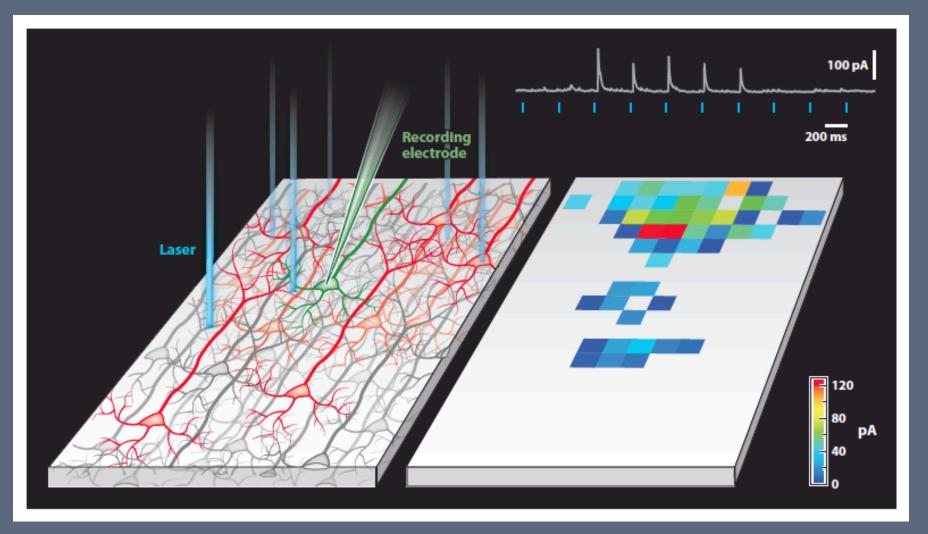


High-speed mapping of synaptic connectivity using photostimulation in Channelrhodopsin-2 transgenic mice

Illumination controls number and frequency of action potentials

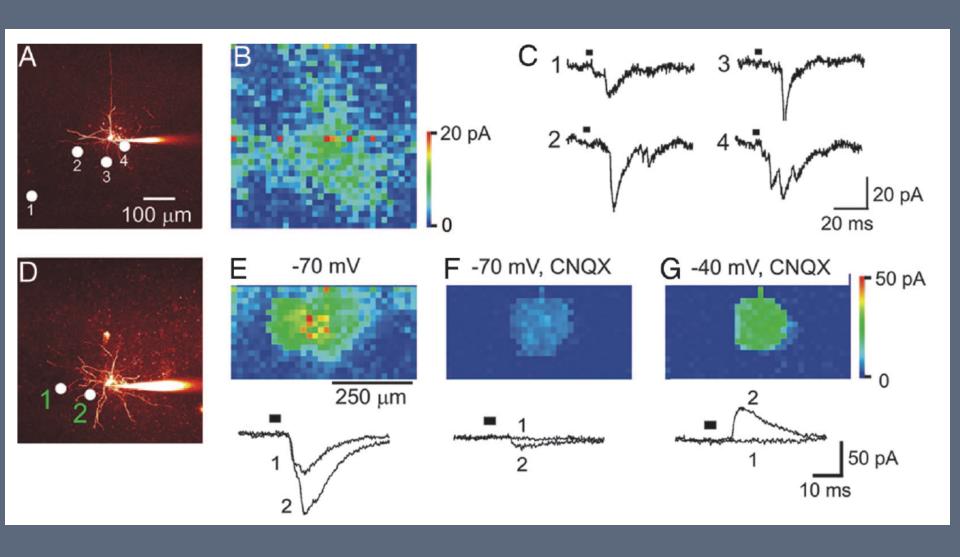


Mapping microcircuits



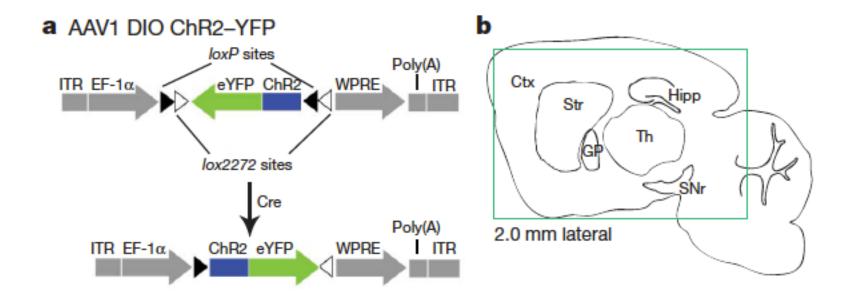
Miesenbock, G. Annual Review of Cell and Developmental Biology (2011)

Properties of cortical microcircuits

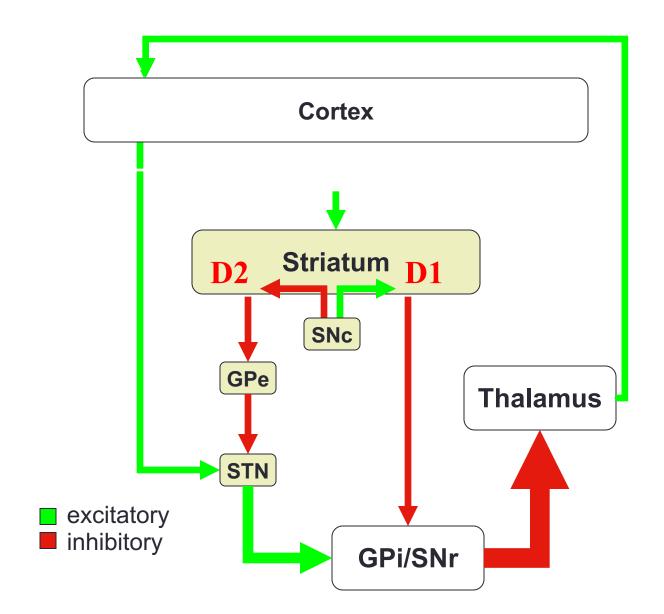


Regulation of parkinsonian motor behaviours by optogenetic control of basal ganglia circuitry

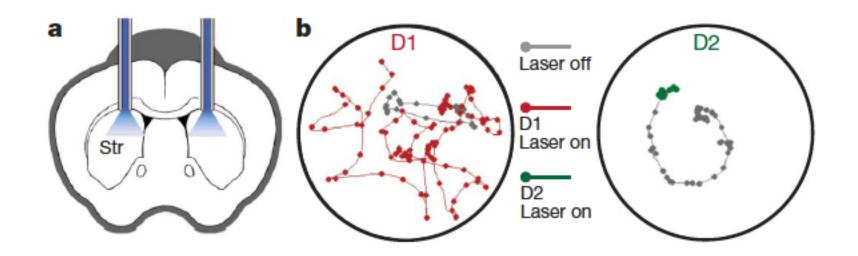
Alexxai V. Kravitz¹, Benjamin S. Freeze^{1,4,5}, Philip R. L. Parker^{1,3}, Kenneth Kay^{1,5}, Myo T. Thwin¹, Karl Deisseroth⁶ & Anatol C. Kreitzer^{1,2,3,4,5}



Cortico-striatal-thalamic-cortical loop



In vivo regulation of motor behavior

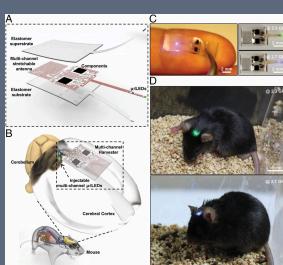


Movie S2: Bilateral illumination of indirect pathway

Kreitzer lab, 2010

Summary

- Optogenetics allows precise control of neuronal circuits
- Several studies have looked at the role of specific brain regions in neuropsychiatric disorders
- It is possible to test predictions on neuronal function
- Optogenetics allows an understanding of the role of specific neuron types
- Limitations:
 - Spatial limitations
 - Invasive
 - Requires gene therapy



OPTOGENETICS